

Experiences with zinc(II) containing artificial metalloproteins and metalloenzymes

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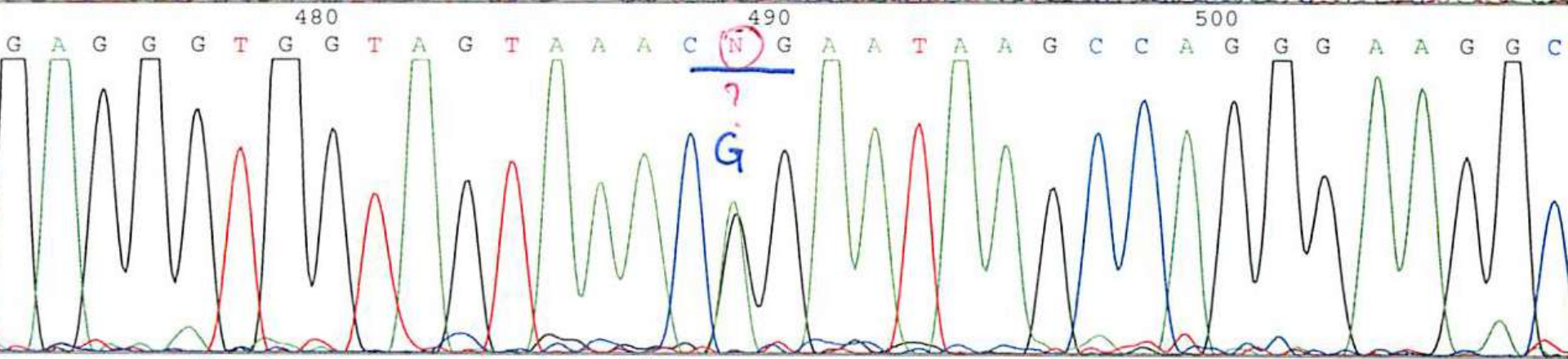
UNIVERSITY OF SZEGED, FACULTY OF SCIENCE AND INFORMATICS

A biologist can do:

- ***Beautiful synthetic reactions: e.g. protein synthesis***
- ***Efficient catalysis: even of the thermodynamically unfavoured reactions – coupling of the reactions by the catalyst***
- ***Extraordinary selectivity: better than the best chemical separation methods.***

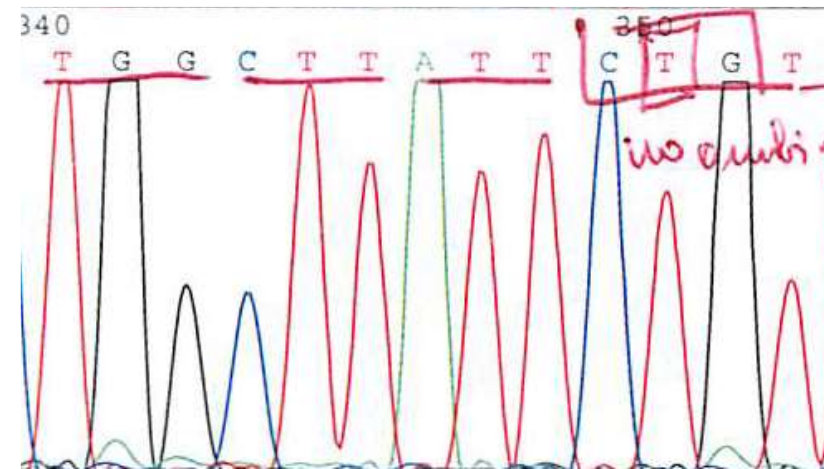
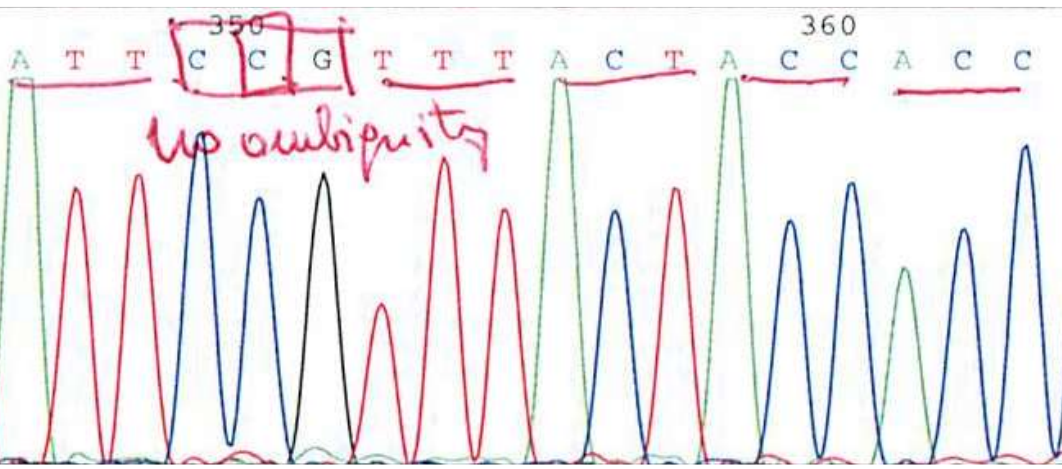
A chemist can do:

Capillary electrophoresis for DNA sequencing

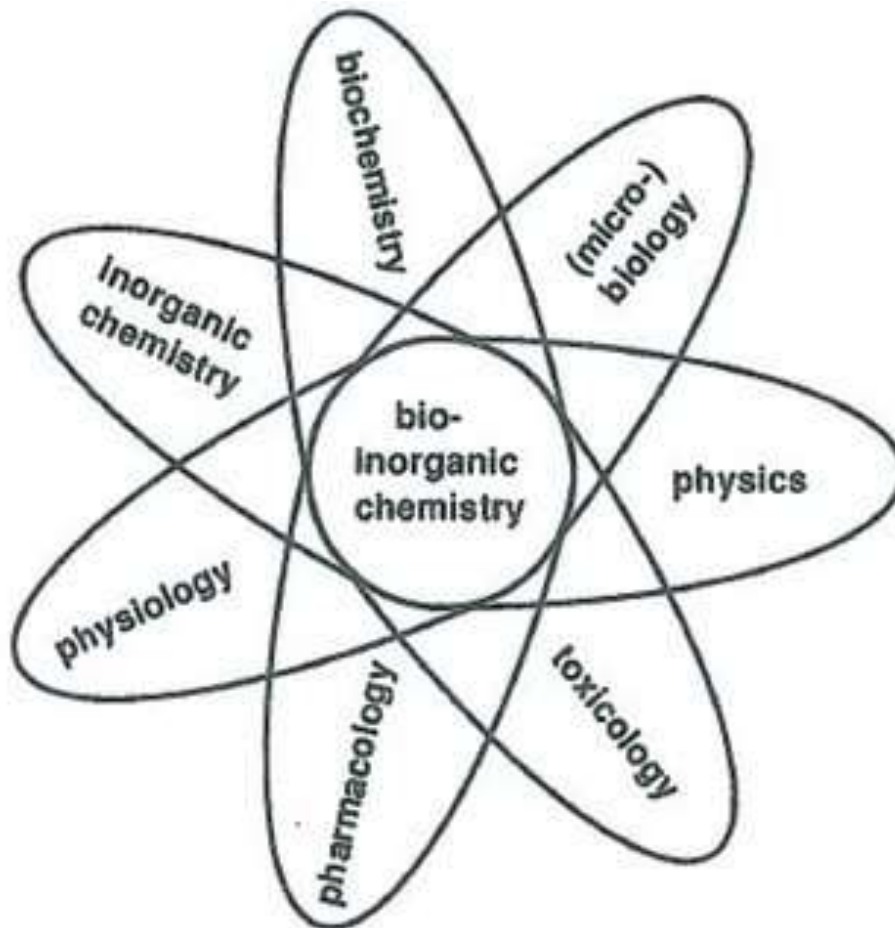


A biologist can do:

Separation of the two large DNA molecules (Same size: ~5000 bp)



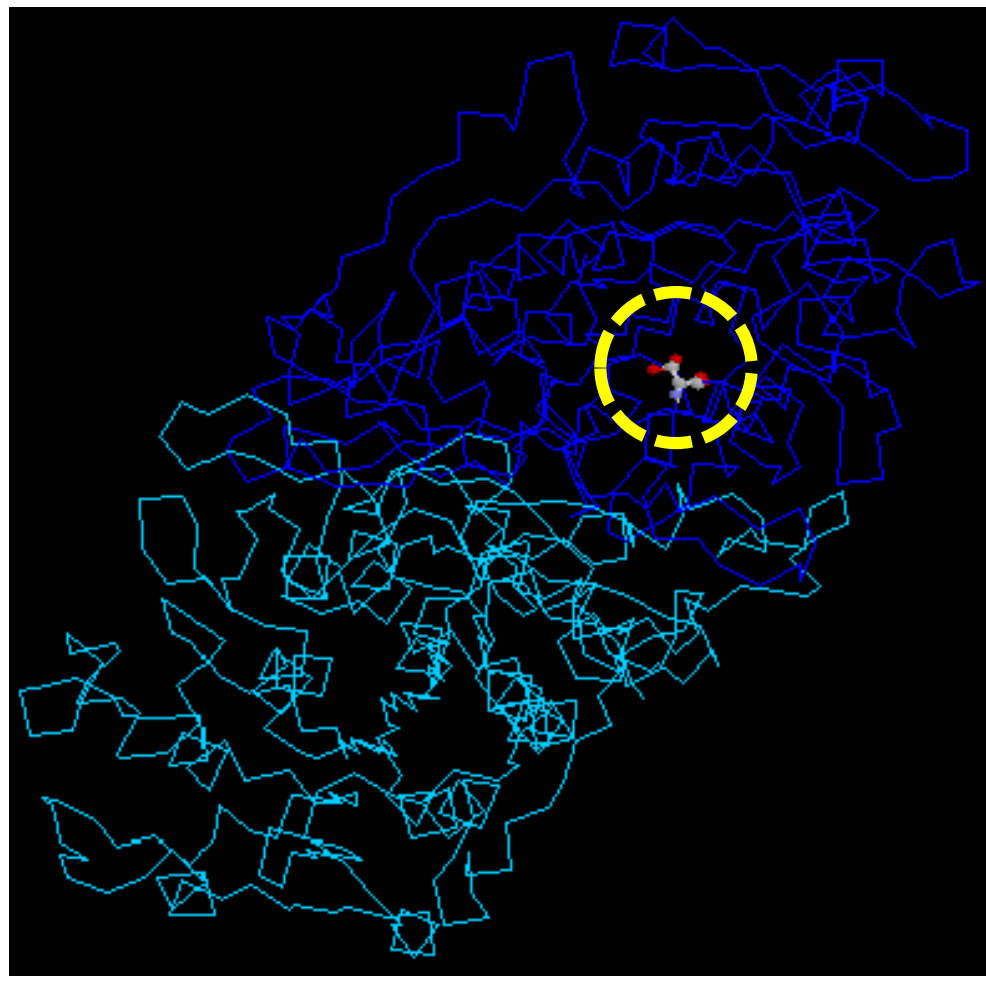
Bioinorganic Chemistry



The role of the metal ions

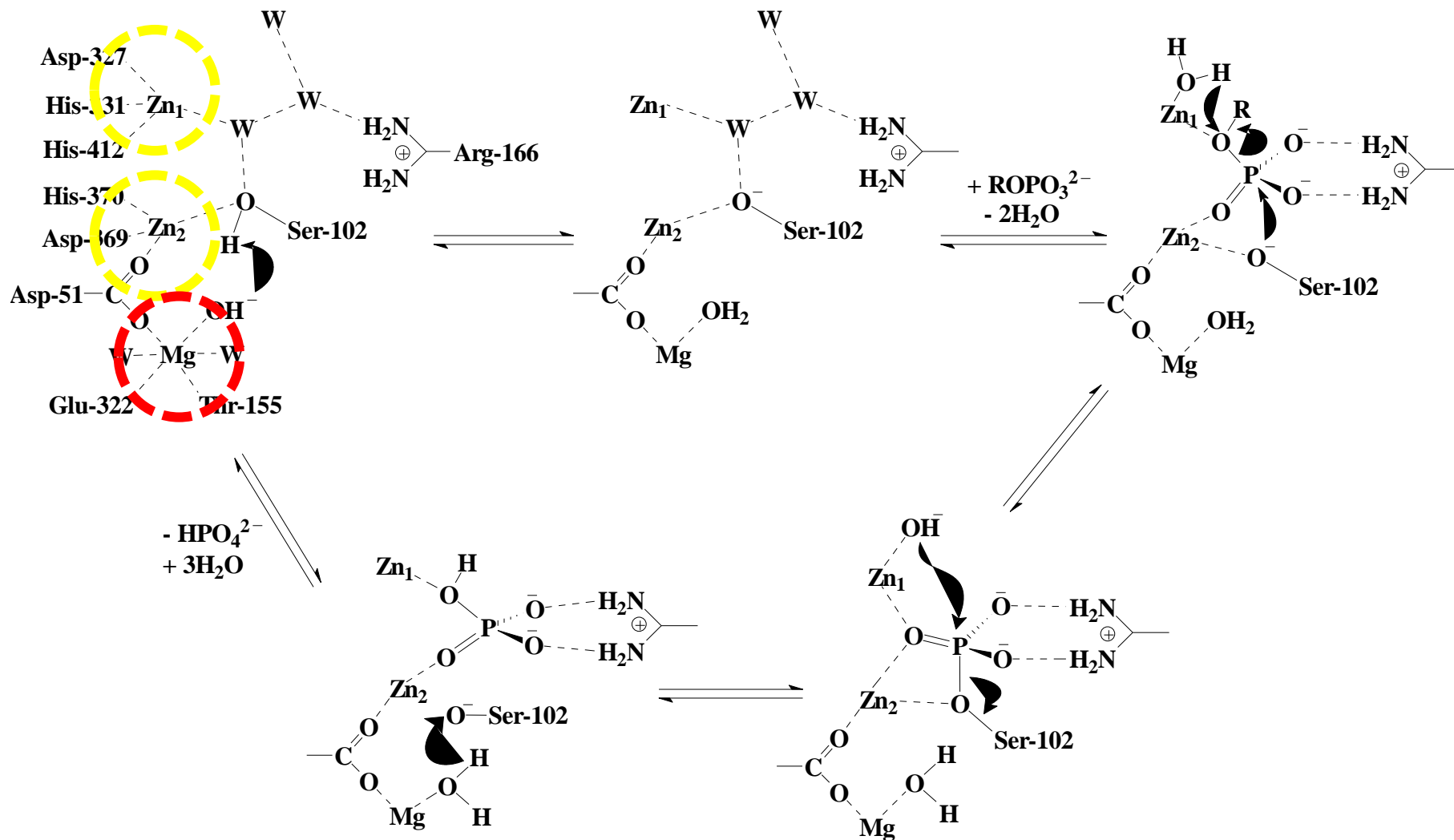


Pdb id: 1ED9



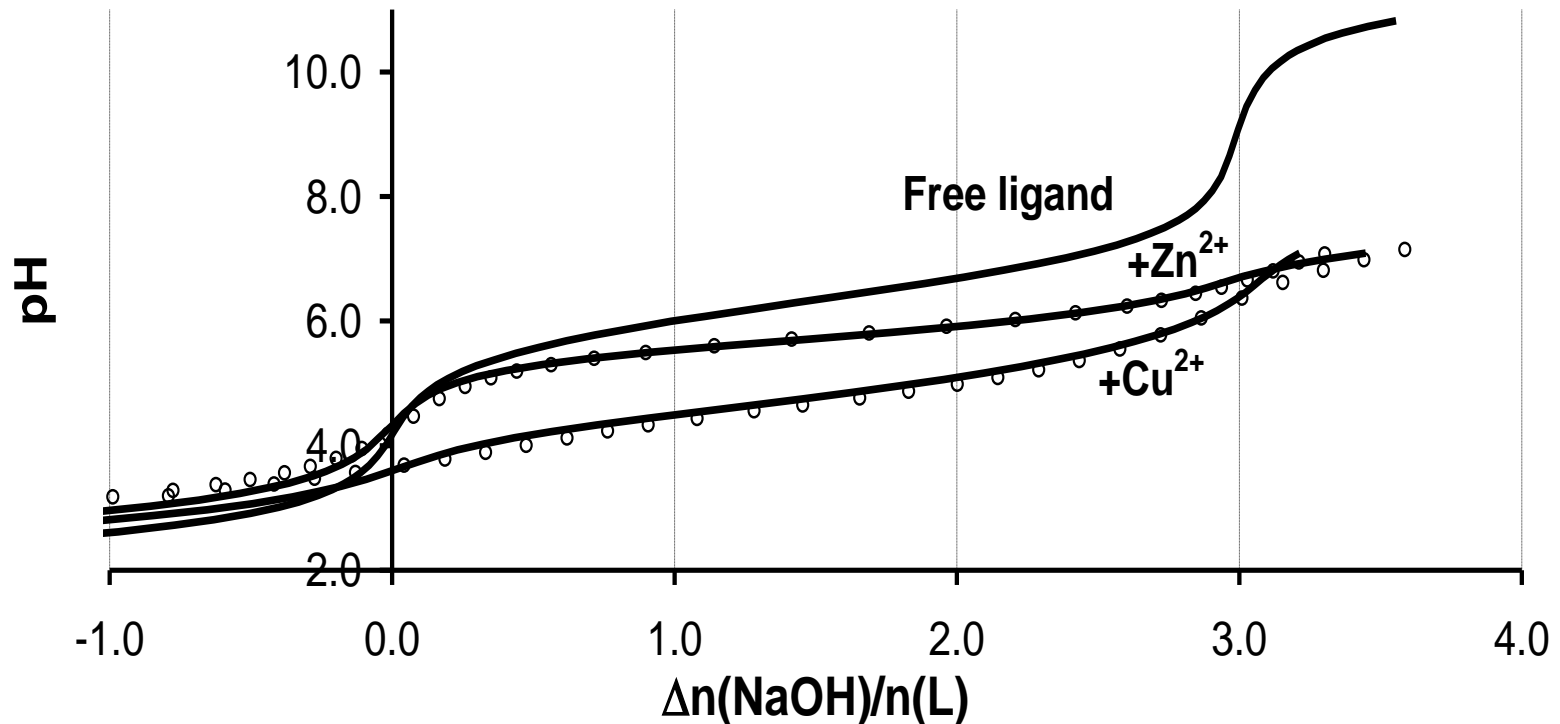
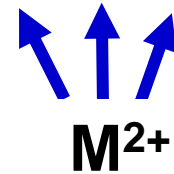
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The role of the metal ions

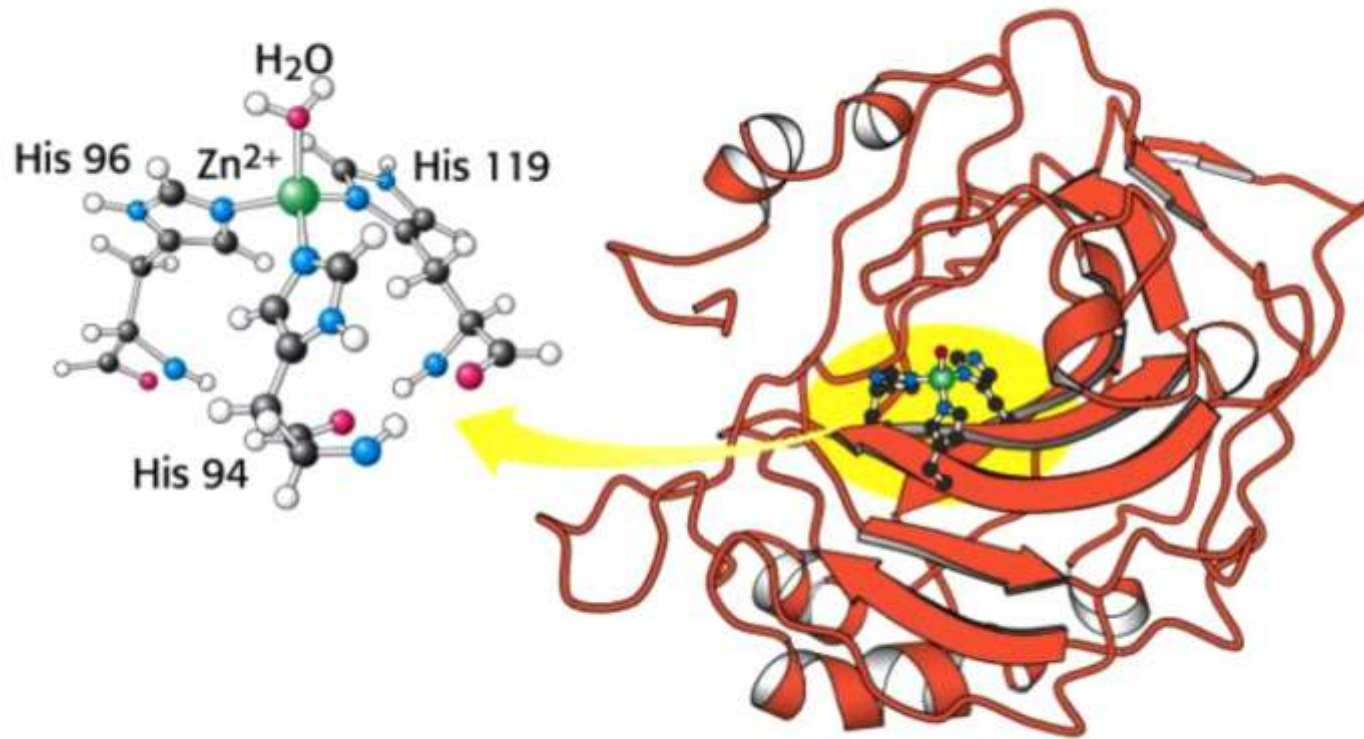


Models vs. Native biomolecules

Histidine triad enzymes: HXHXH motif
Copper(II) and zinc(II) complexes of the AcHPHPH-CONH₂ peptide



Models vs. Native biomolecules



Bioinorganic zinc chemistry

$\beta\beta\alpha$ -metal ion binding motifs:

Cys_2His_2 zinc finger

vs.

HNH:

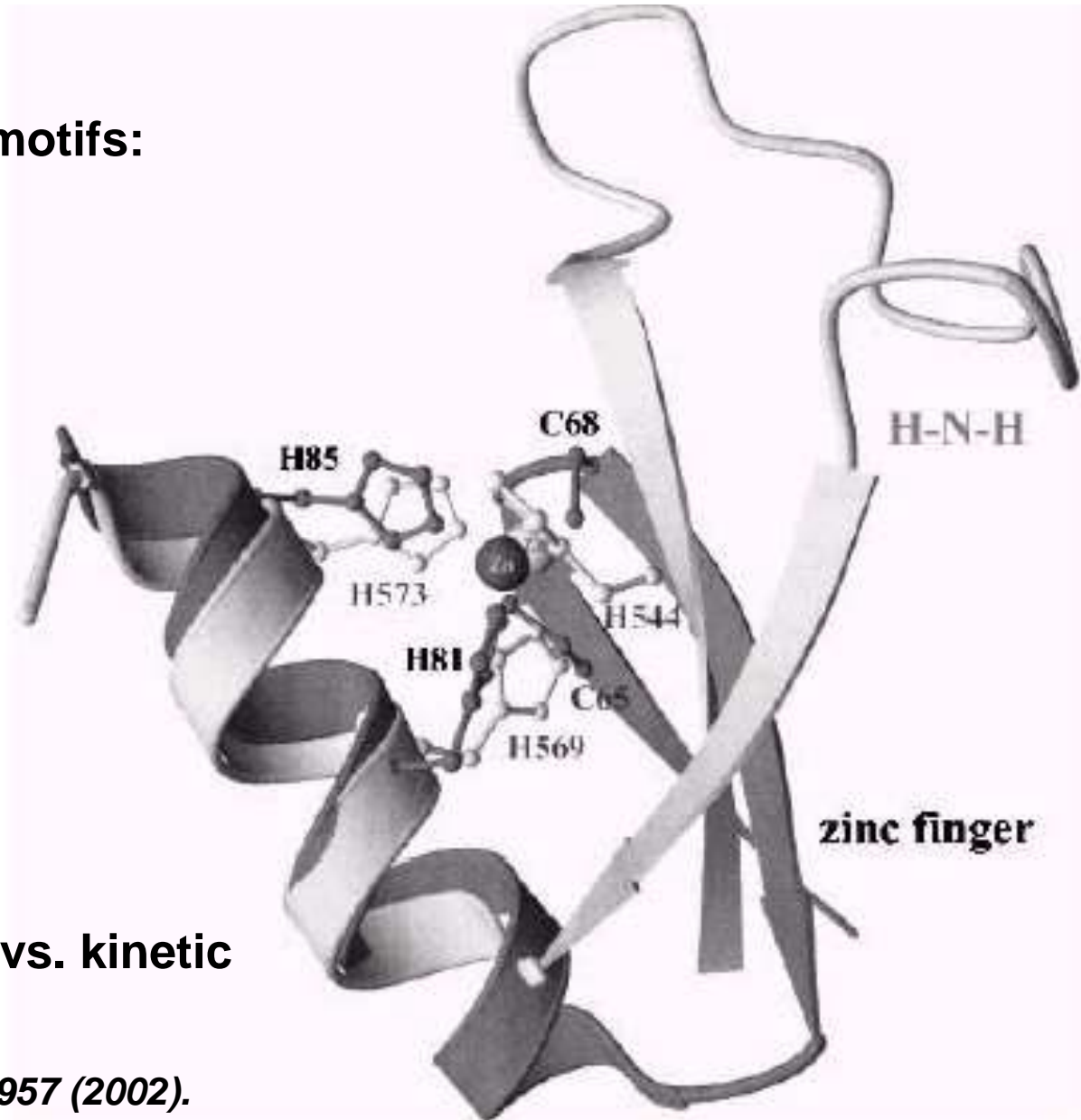
$\text{HHX}_{14}\text{NX}_8\text{HX}_3\text{H}$

Redox properties

Role of metal ion

Geometry

Stability – thermodynamic vs. kinetic





Chris Oostenbrink Sine Larsen Lars Hemmingsen Peter Thulstrup Hans Christensen Masamitsu Asaka Kyosuke Nagata Wojciech Bal



http://www2.sci.u-szeged.hu/artmetprot/index_eng.htm

Artificial nucleases

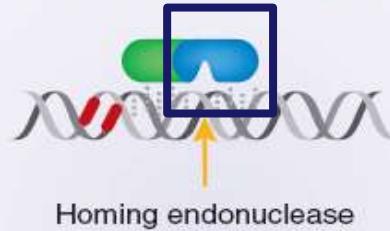
DNA containing mutated sequence



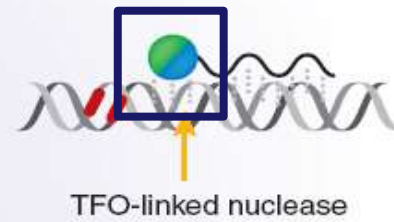
Add cleavage reagent and corrected DNA template



Zinc-finger nuclease



Homing endonuclease

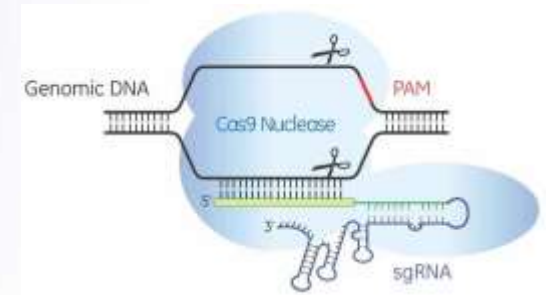


TFO-linked nuclease

Homology-directed repair



Corrected gene



CRISPR/Cas9

Non-regulated
nuclease domain

© 2007 Caesar

Artificial nucleases



Artificial nucleases

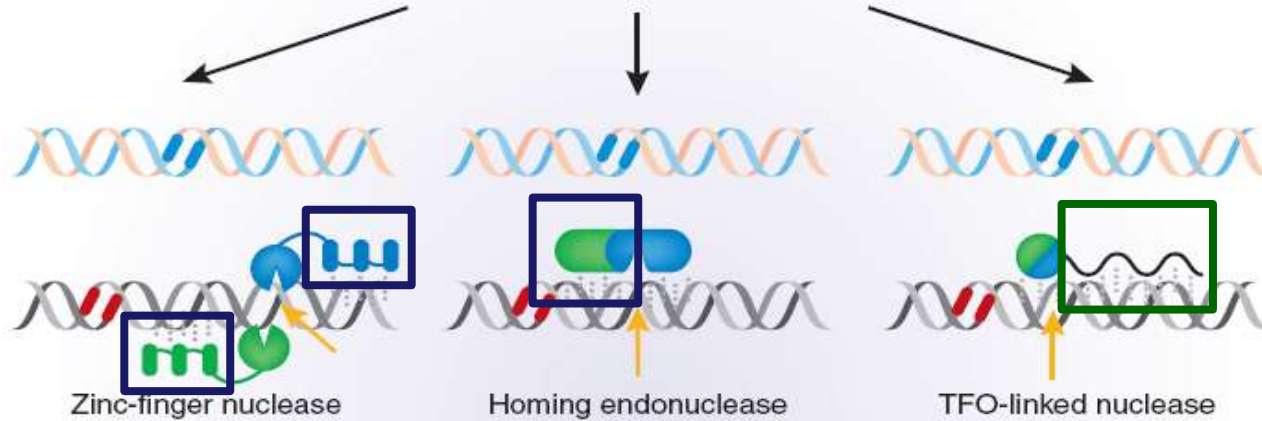


Artificial nucleases

DNA containing mutated sequence

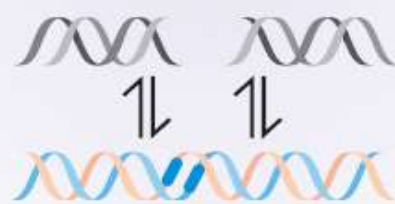


Add cleavage reagent and corrected DNA template



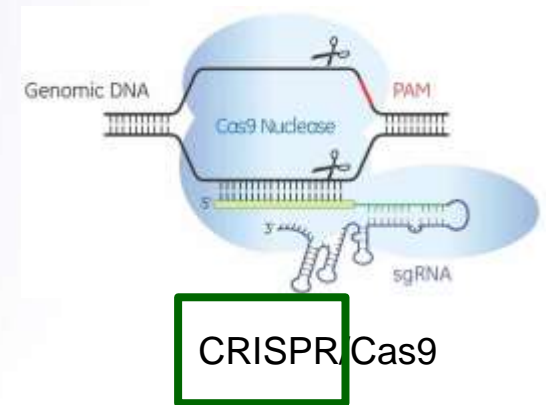
Protein guide

Homology-directed repair



Corrected gene

RNA guide



from Caesar

Artificial nucleases

A guide to genome engineering with programmable nucleases

Table 2 | Comparison of three classes of programmable nucleases*

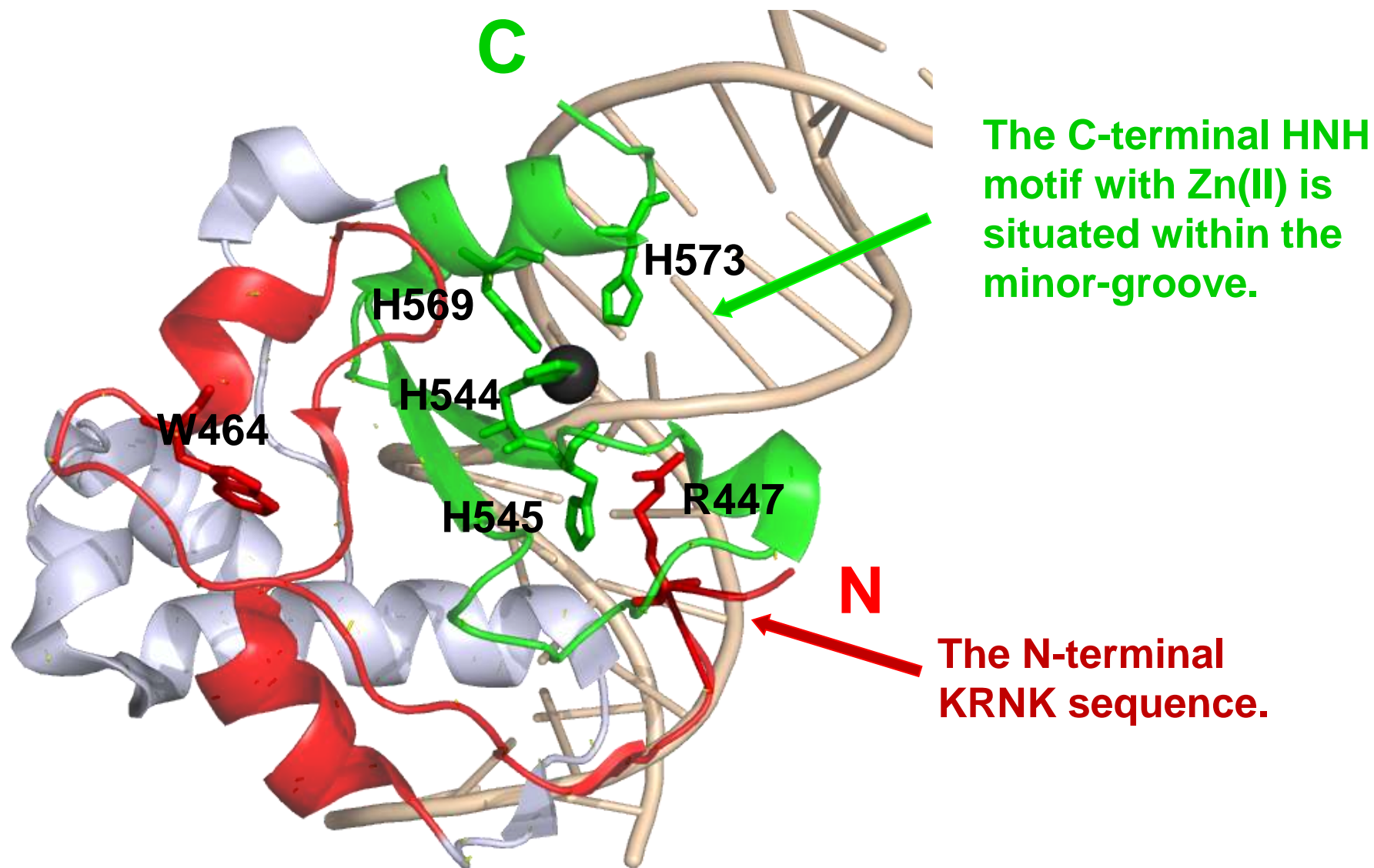
	ZFNs	TALENs	RGENs
DNA targeting specificity determinant	Zinc-finger proteins	Transcription activator-like effectors	crRNA or sgRNA
Nuclease	<i>FokI</i>	<i>FokI</i>	Cas9
Success rate[‡]	Low (~24%)	High (>99%)	High (~90%)
Average mutation rate[§]	Low or variable (~10%)	High (~20%)	High (~20%)
Specificity-determining length of target site	18–36 bp	30–40 bp	22 bp (total length 23 bp)
Restriction in target site	G-rich	Start with T and end with A (owing to the heterodimer structure)	End with an NGG or NAG (lower activity) sequence (that is, PAM)
Design density	One per ~100 bp	At least one per base pair	One per 8 bp (NGG PAM) or 4 bp (NGG and NAG PAM)
Off-target effects	High	Low	Variable
Cytotoxicity	Variable to high	Low	Low
Size	~1 kb×2	~3 kb×2	4.2 kb (Cas9 from <i>Streptococcus pyogenes</i>) + 0.1 kb (sgRNA)

Aim of the work

To develop an artificial nuclease with a safe control mechanism:

The nuclease has to stop cleaving DNA in case of any damage of the enzyme within the cells.

Crystal structure of an NCoIE7 mutant in its DNA complex

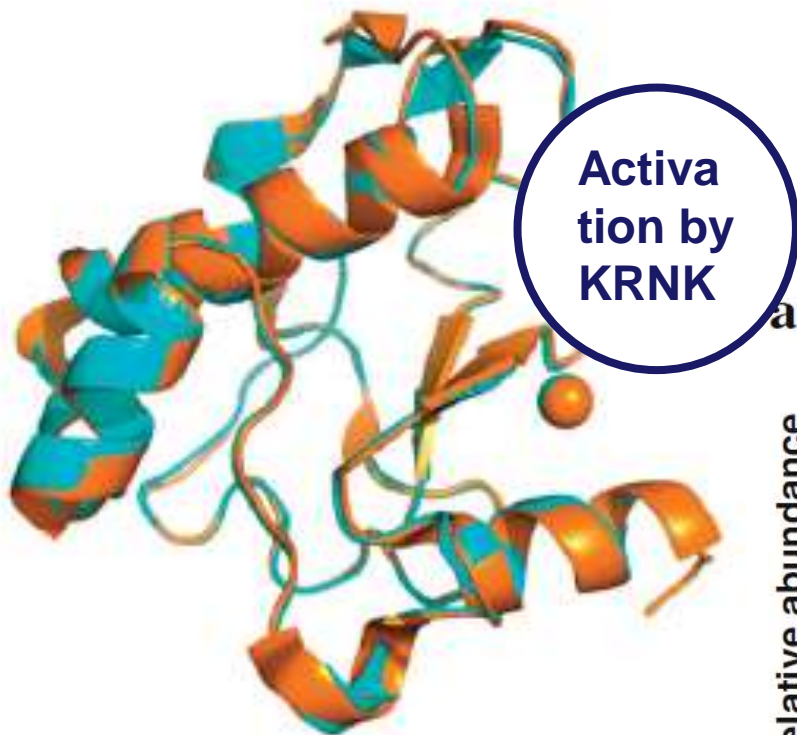


Y.-T. Wang, J.D. Wright, L.G. Doudeva, H.-C. Jhang, C. Lim, H.S. Yuan, J. Am. Chem. Soc. 131 (2009) 17345-17353; Protein Data Bank (PDB) ID: 3FBD

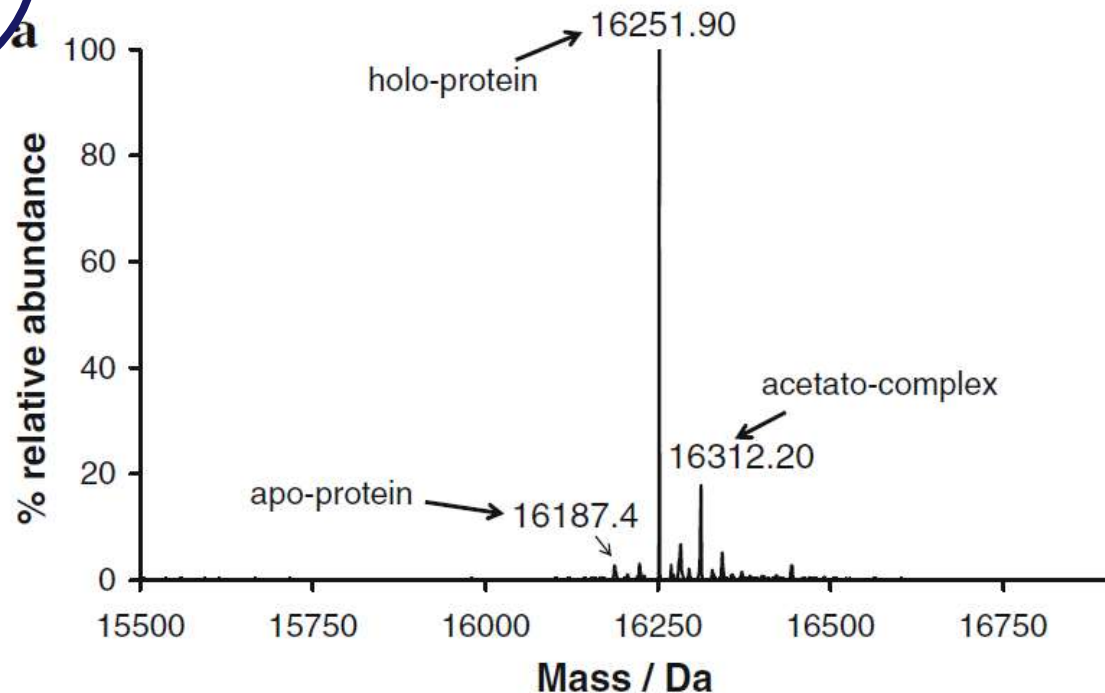
NCoIE7

- 1. Interaction between the N- and C-termini***
- 2. Fine tuning of the nuclease activity***
- 3. W464: stabilization of the structure of NCoIE7***
- 4. Inducible preorganized zinc(II)-binding site***

1. Interactions between the N- and C-termini

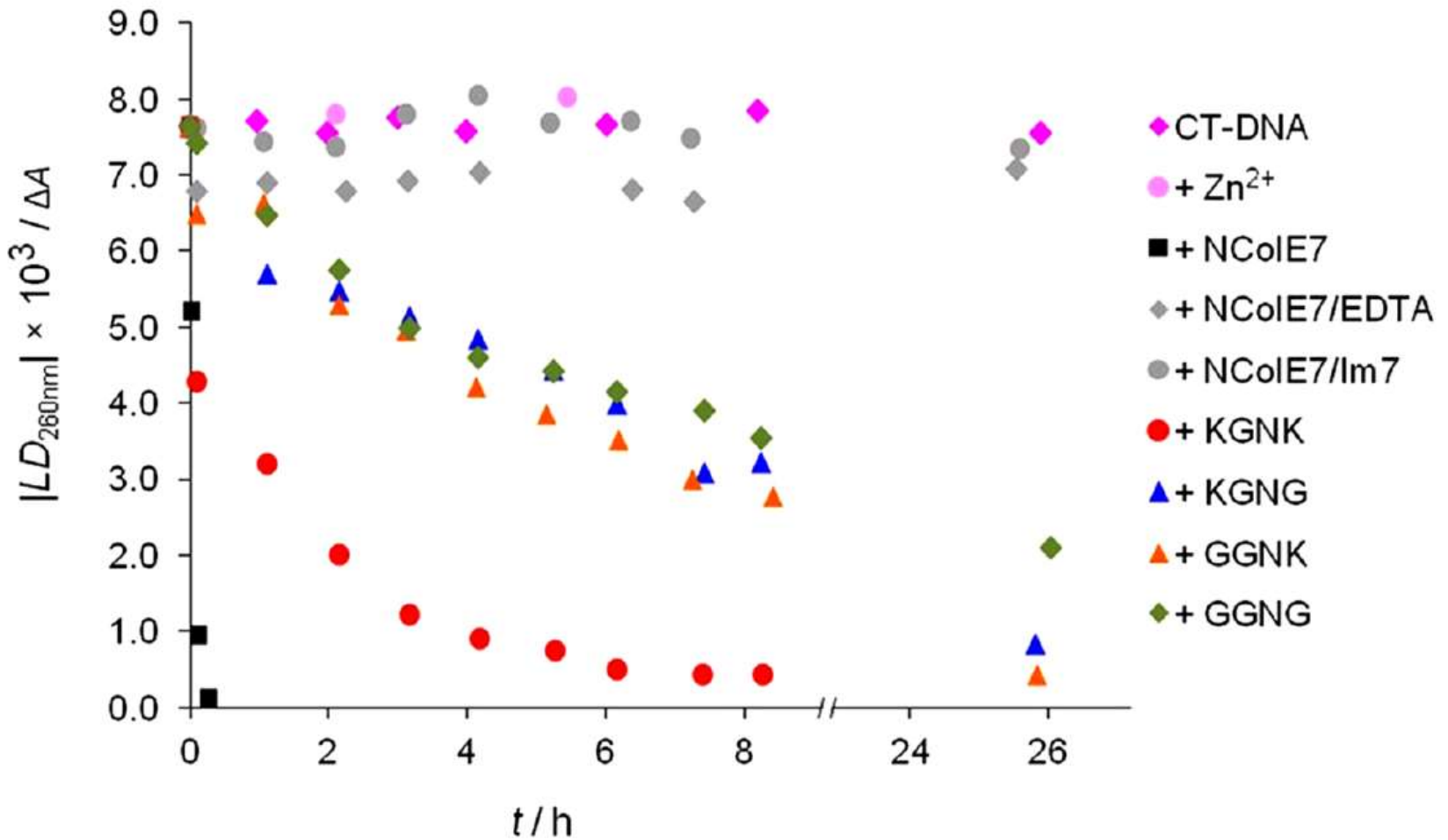


Δ N4-NC_oIE7(-C*):
~~KRNK~~

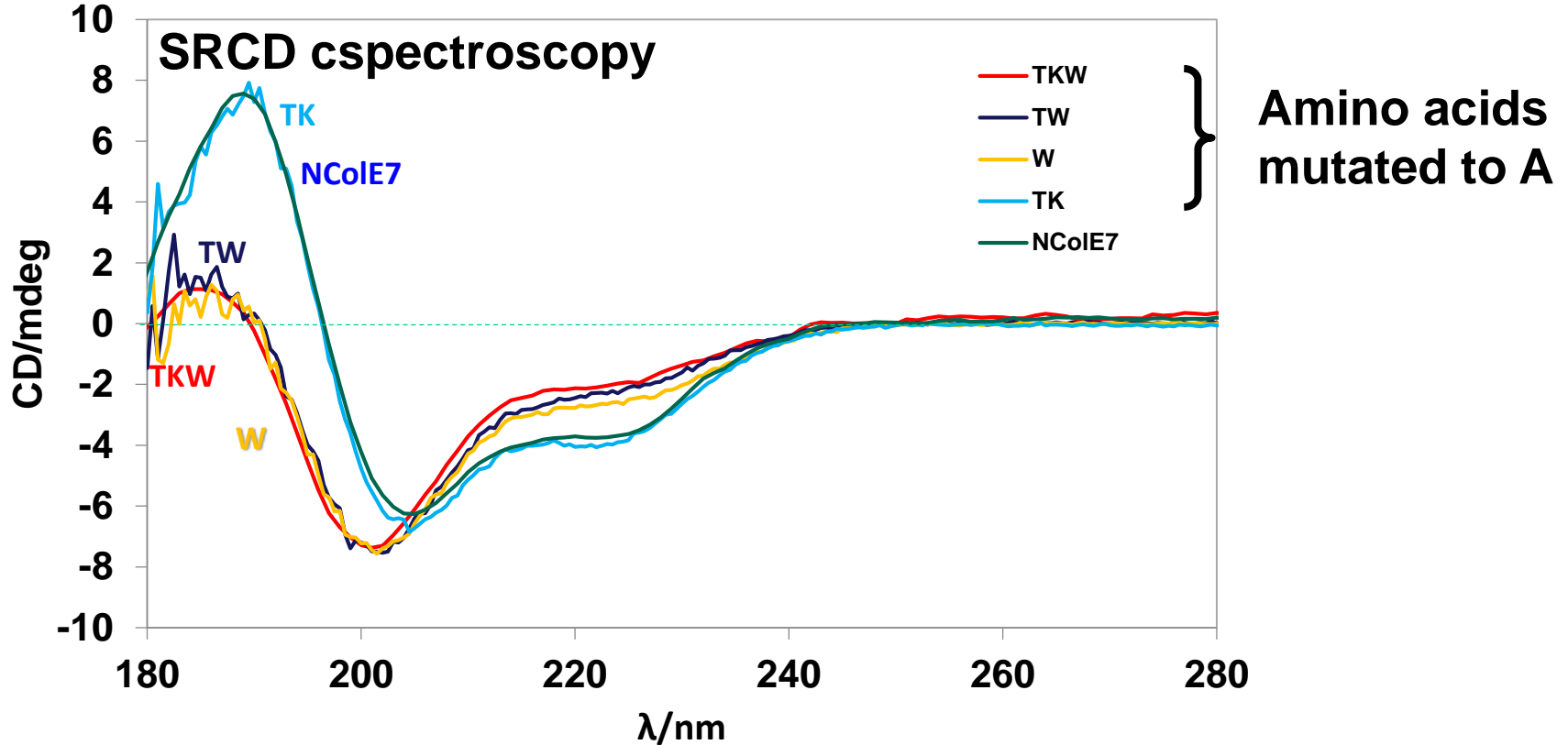


Gyurcsik, et al., *J. Biol. Inorg. Chem.*, 18, 309-321 (2013); *Acta Cryst. Sect F.*, 69 (2013) 551-554; *Metallomics*, 6 (2014) 2090-2099.

2. Fine tuning of the nuclease activity



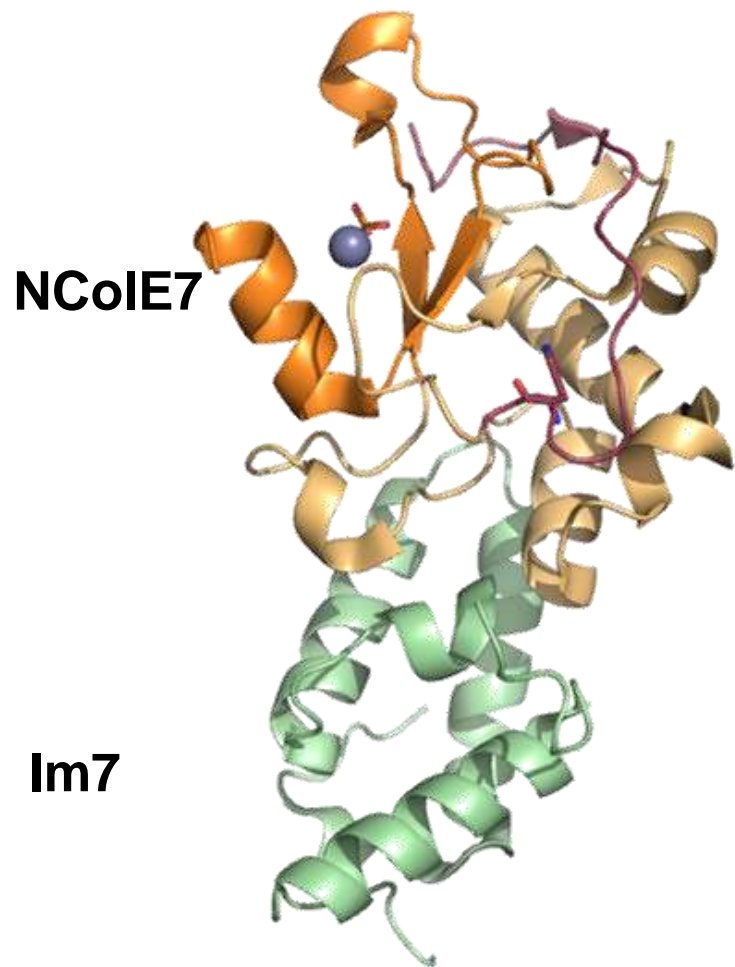
3. W464: stabilization of the structure of NCoIE7



KRNKPGKATGKGPVNNK**W**LNNAGKDLGSPVPDRIANKLRDKEF
 KSFDDFRKKFWEEVSKDPELSKQFSRNNNDRMKVGKAPKTRTQD
 VSGKRTSFELHHEKPISQNGGVYDMDNISVVTPKRHIDIHRGK

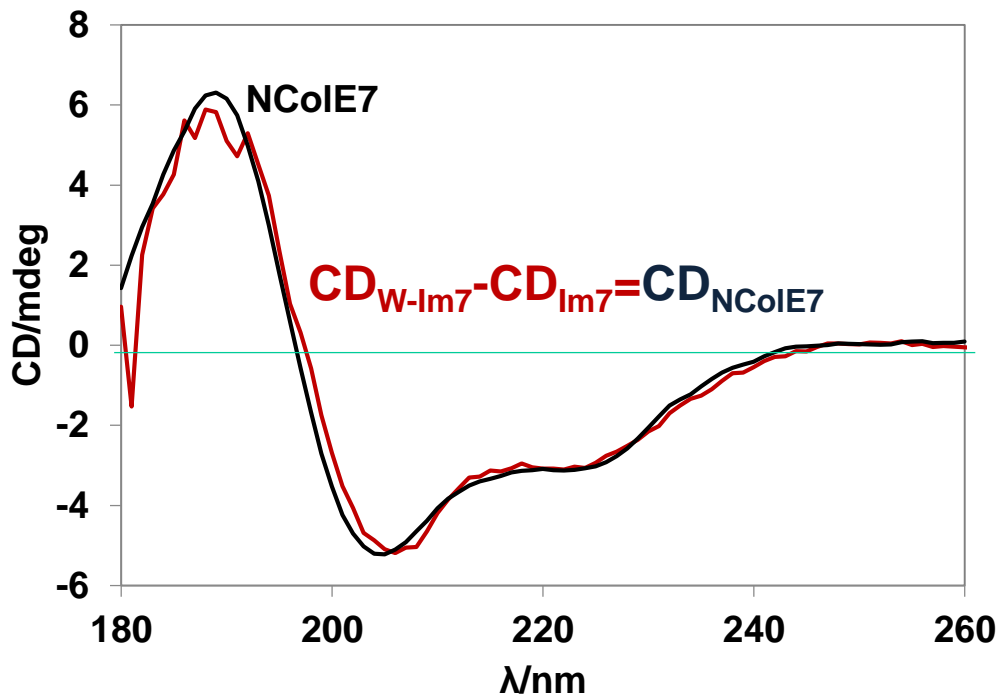
Gyurcsik, et al., J. Biol. Inorg. Chem., 19 (2014) 1295-1303; Gyurcsik, Curr. Prot. Pept. Sci., 17 (2016) 191-197; Gyurcsik, et al., J. Inorg. Biochem. 151 (2015) 143-149; Protein Sci., 25 (2016) 1977-1988.

4. Inducible preorganized zinc(II)-binding site



Inducible preorganized metal ion binding site

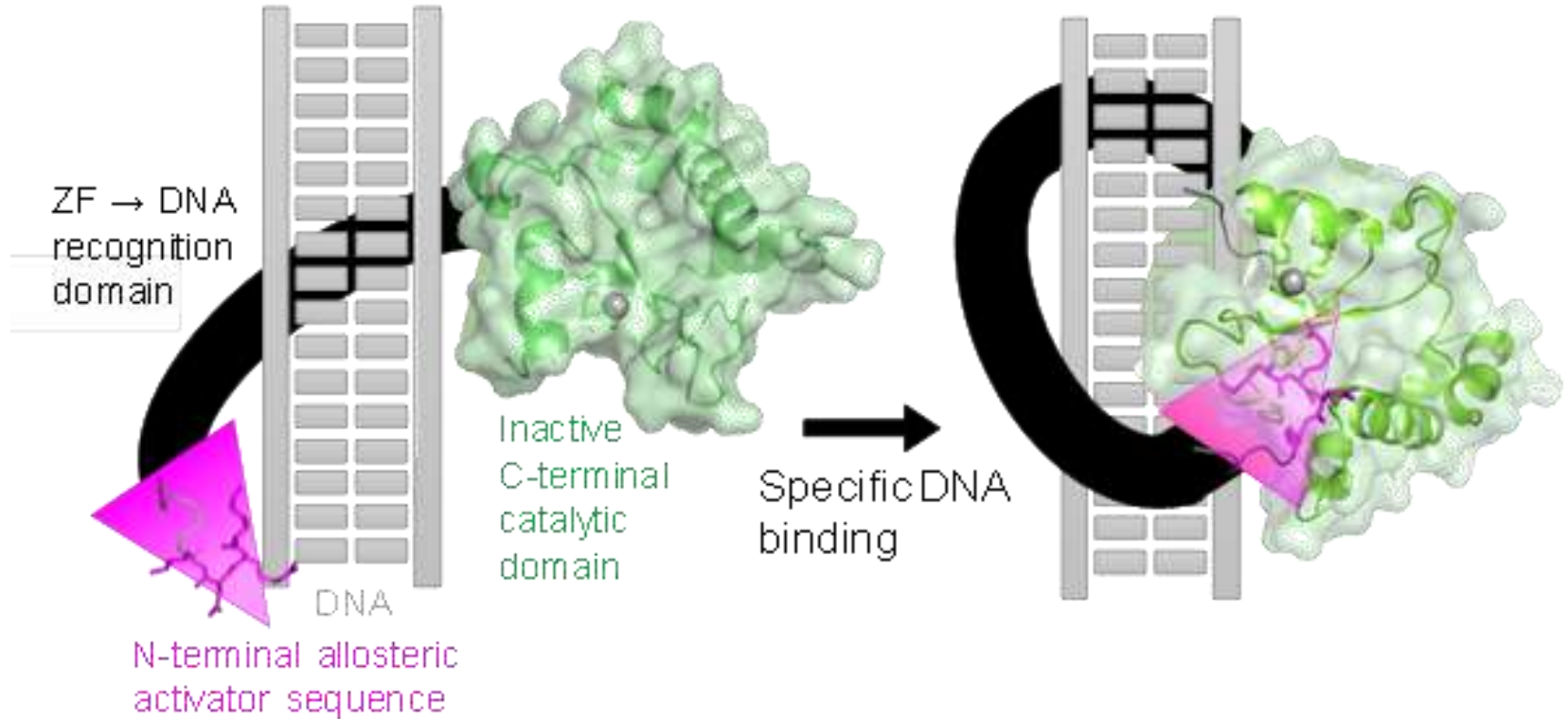
SRCD spectroscopy



Isothermal Titration Microcalorimetry Zn(II) binding

	K_d (Prot+Im7)	K_d (Prot)
NCoIE7	61 ± 18 nM	9.6 ± 3.2 nM
TKW	33 ± 23 nM	11 ± 1 μM
W	55 ± 25 nM	5.6 ± 0.3 μM
Im7	ND	

The concept of the NCoIE7-based artificial nuclease design



Split NCoIE7 into **Nx** regulatory and **Cy** catalytic segments and insert a zinc finger guide protein between them.

N-terminal control unit of NCoIE7

Linker

Zinc Finger protein

Linker

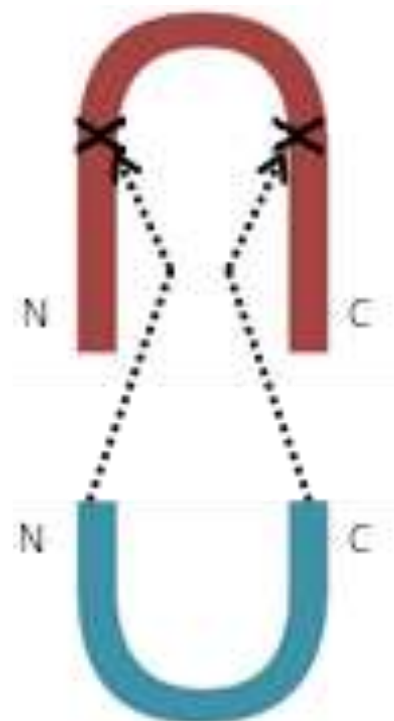
C terminal catalytic unit of NCoIE7

Protein design

Strategy:

Nx – **ZF** – **Cy**

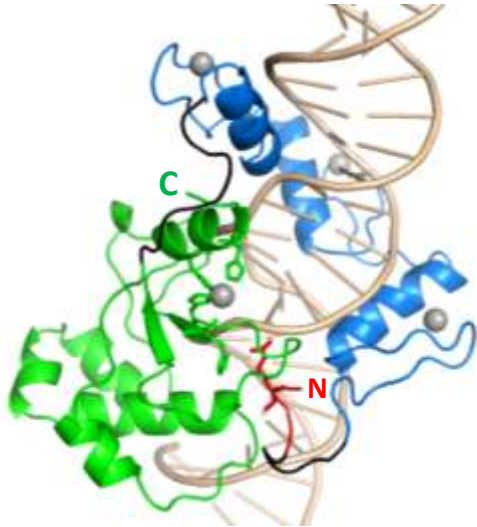
NCoIE7



Zinc finger

Nx – ZF – Cy strategy

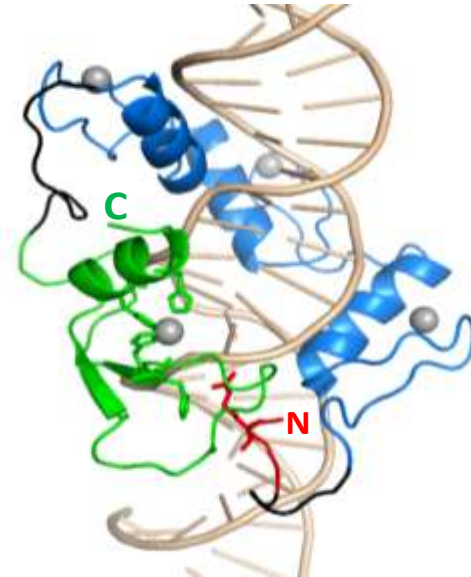
Protein expression and purification



N4-ZF-C105



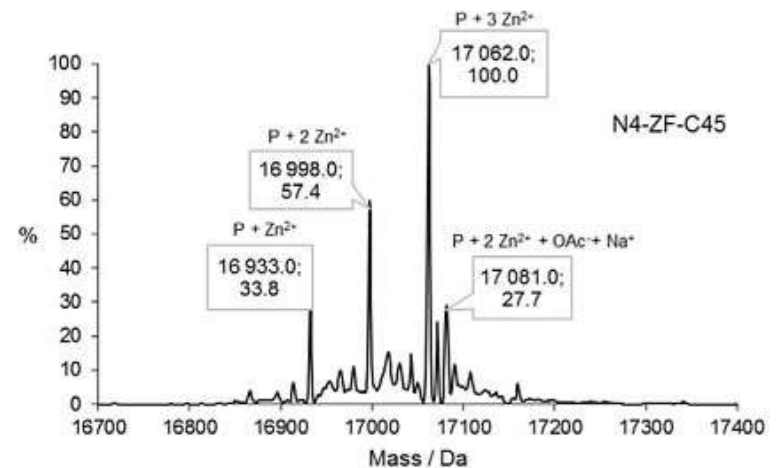
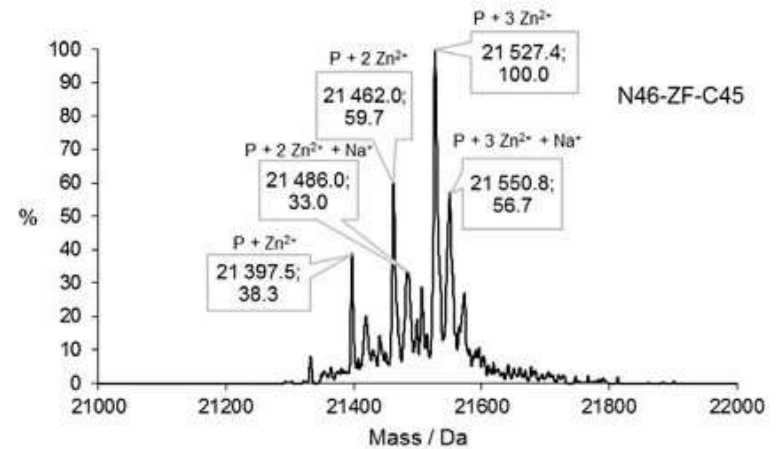
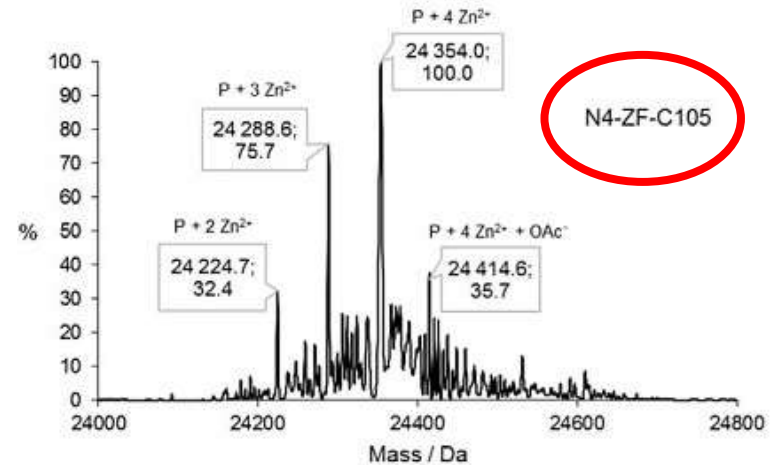
N46-ZF-C45



N4-ZF-C45

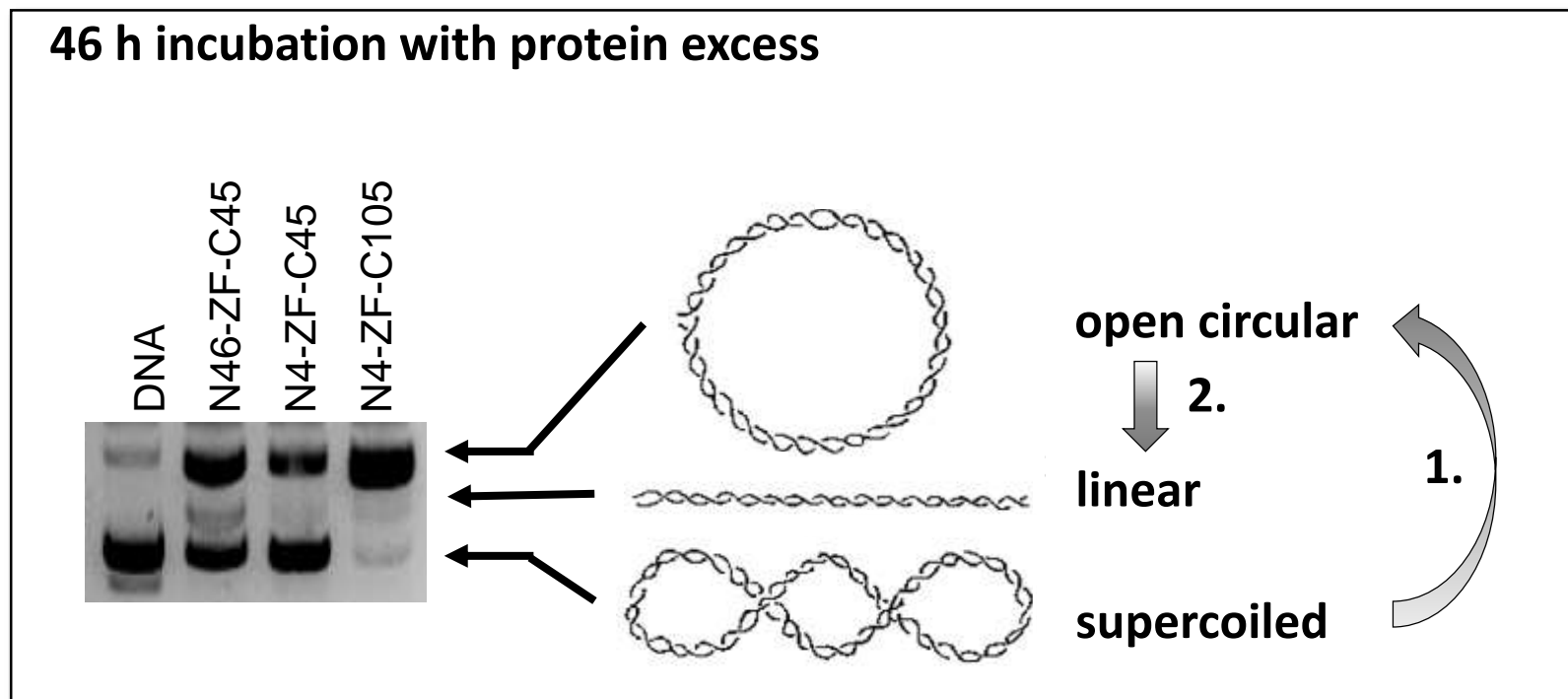
Nx – ZF – Cy strategy

Protein expression and purification



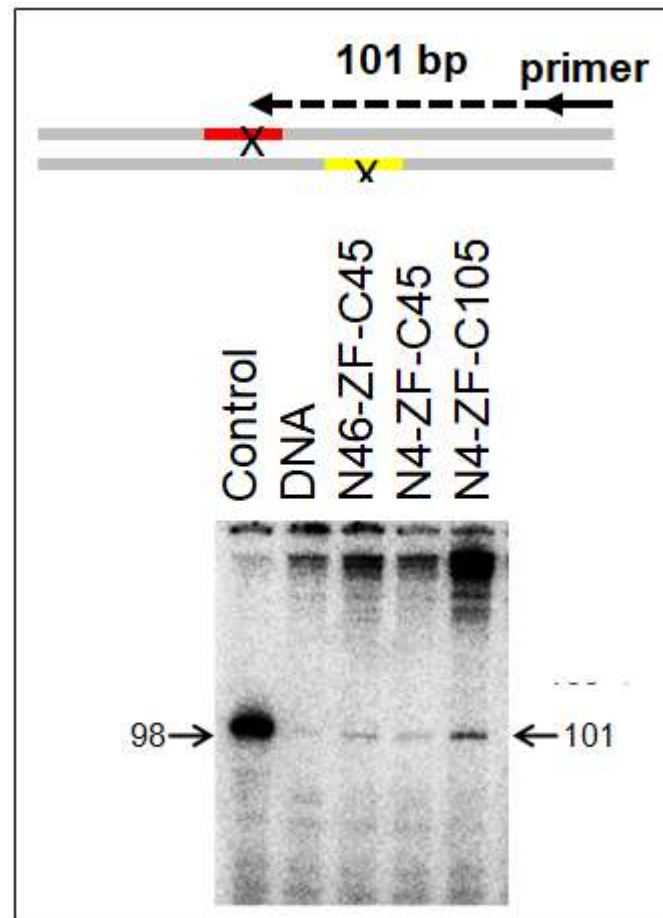
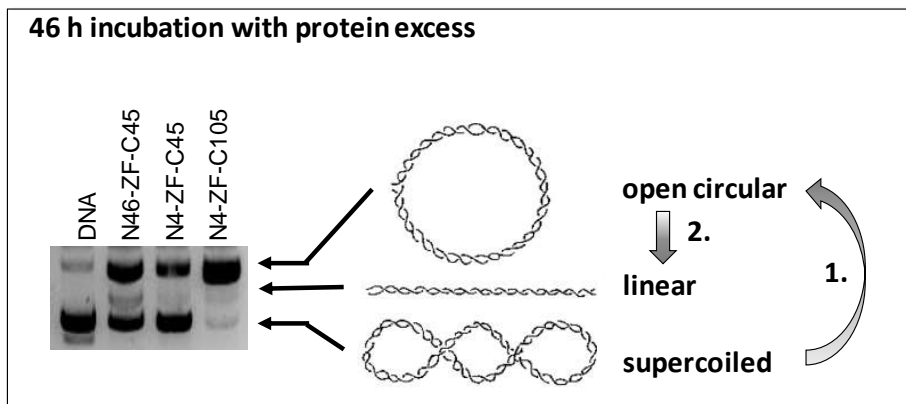
Nx – ZF – Cy strategy

Catalytic activity - specificity



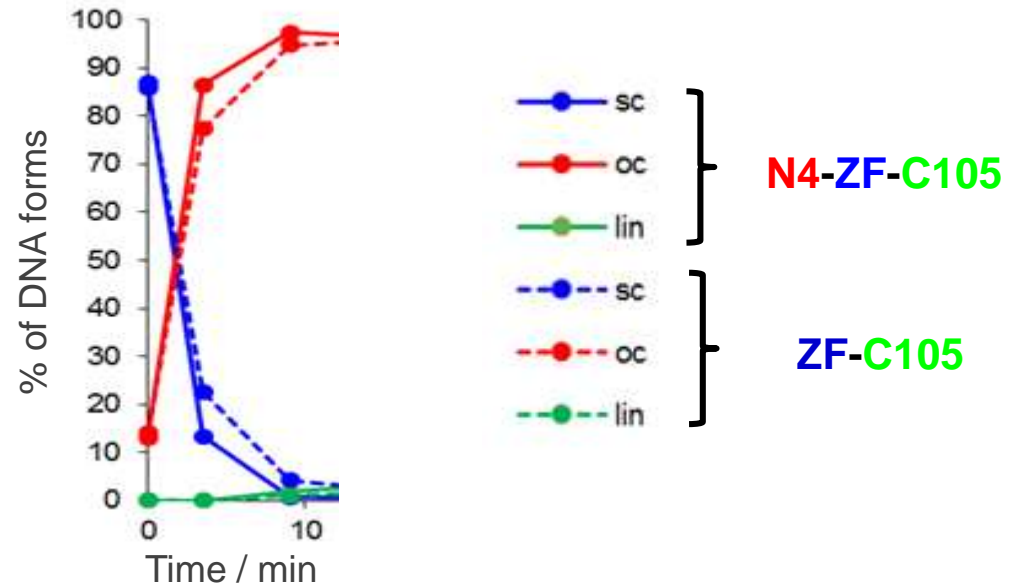
Nx – ZF – Cy strategy

Catalytic activity - specificity

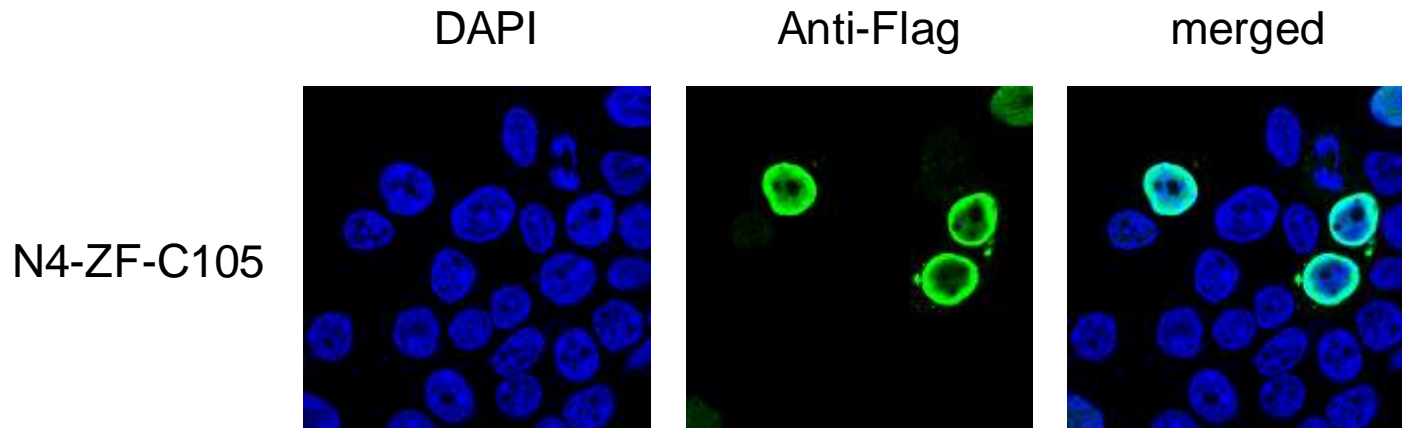


Nx – ZF – Cy strategy

Allosteric control



Cellular localization on in HEK293T cells



Conclusions

- *The formation of native-like structure of the artificial nucleases can be monitored through their zinc-binding ability.*
- *The Nx – ZF – Cy models showed low activity, they were specific, but not well regulated.*
- *The possibility of fine tuning and structure induction in combination with competition by the DNA binding proteins will lead us to a better artificial enzyme.*

Acknowledgement



This work was financially supported by the research grants

NKFIH K_16/120130 and

GINOP 2.3.2-15-2016-00038.



PROJECT FINANCED
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